



**VASOACTIVE EICOSANOIDS:
SYNTHESIS OF 12,20-DIHYDROXYEICOSA-5(Z),8(Z),10(E),14(Z)-
TETRAENOIC ACID VIA A NOVEL CHIRAL BIS-LACTOL**

J. R. Falck,*^{1,2} Kamlesh Chauhan,¹ Mark Rosolowsky,² and William B. Campbell²

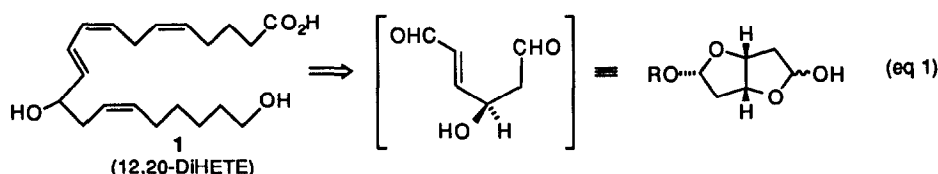
Departments of Molecular Genetics¹ and Pharmacology²

University of Texas Southwestern Medical Center

Dallas, Texas 75235

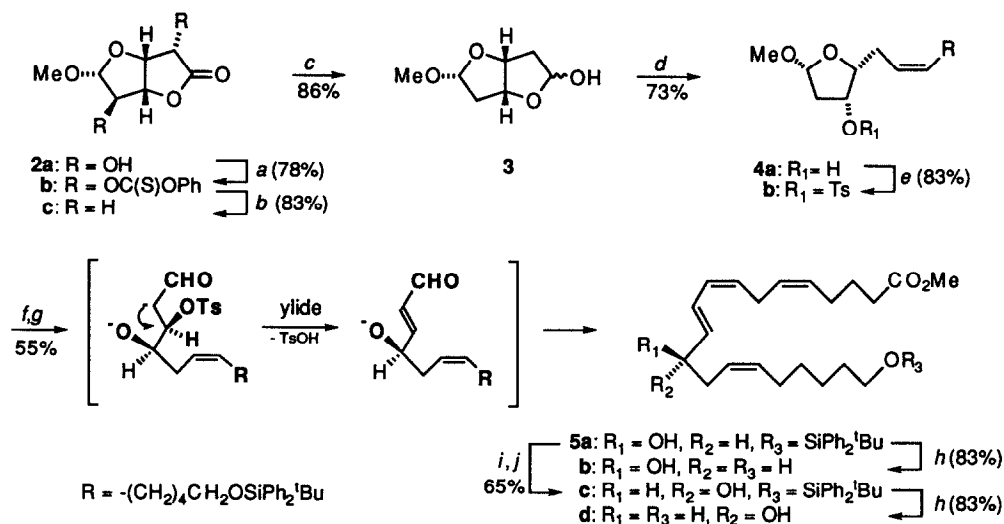
Abstract: The type IIB eicosanoids, 12(R),20- and 12(S),20-dihydroxyeicosatetraenoic acid (DiHETE), were conveniently synthesized utilizing a carbohydrate-derived bis-lactol as a differentiated 4-hydroxyhex-2(E)-enedial chiron. 12(S),20-DiHETE, but not the 12(R)-isomer, induced vasodilation in precontracted canine arteries.

The biogenesis of 12-hydroxyeicosatetraenoic acid (12-HETE) in mammals presents a unique paradigm, i.e., arachidonic acid is converted to either 12(S)-HETE by lipoxygenase^{1,2} or to its R-antipode by cytochrome P450.³ Depending on the tissue, both metabolic routes can function concurrently. The enantiomers display qualitatively and quantitatively different pharmacologic profiles,⁴ yet both can be converted to 12,20-DiHETE⁵ (1) via type IIB⁶ transcellular metabolism. Compared to 12-HETE, relatively little is known about the physiological role(s) and the absolute configuration of endogenous 1. Herein, we describe a convenient synthesis⁷ of 12(S),20- and 12(R),20-DiHETE utilizing a carbohydrate-derived bis-lactol as a differentiated 4-hydroxyhex-2(E)-enedial chiron (eq 1) and describe the initial biological evaluation of both antipodes in canine coronary artery.



Methyl β -furanoside **2a**, readily obtained⁸ from commercial D-glucurono-6,3-lactone in 80% yield, was smoothly deoxygenated by way of its 2,5-bis(phenylthionocarbonate) **2b** (mp 164-66°C) using Bu_3SnH in toluene (Scheme 1).⁹ Subsequent low temperature diisobutylaluminum hydride (DIBAL-H) reduction of the resultant lactone **2c**¹⁰ (mp 103-4°C) gave rise to the strategic chiral bis-lactol **3**, whose implicit aldehydes were accessed individually. The first was homologated using 6-(*tert*-butyldiphenylsilyloxy)hexyldienetriphenylphosphorane¹¹ (**6**) leading to furanoside **4a**. Following tosylation of the newly exposed secondary alcohol, the remaining lactol was unlocked by mild acidic hydrolysis. Final elaboration to complete the eicosanoid carbon skeleton, thus furnishing mono-protected diol **5a**, exploited the rapid, ylide-induced elimination of tosylate from the open-chain tautomer of **4b**.¹² This generated a hydroxy-

Scheme 1



^aPhOC(S)-Cl, N-hydroxysuccinimide (5 mol %), C₅H₅N/CH₃CN, 60°C, 5 h. ^bBu₃SnH, AIBN, PhCH₃, 80°C, 3 h. ^cDIBAL-H, PhCH₃, -78 to 0°C, 2 h. ^d6 (3 equiv), THF/HMPA/PhCH₃ (1:0.8:4), -78 to -20°C, 4 h. ^eTsCl, C₅H₅N/CH₂Cl₂ (1:1), 0 to 23°C, 8 h. ^fAcOH/THF/H₂O (2:1:1), 45°C, 13 h. ^g7 (3.3 equiv), THF/HMPA (10:1), -20°C, 1 h. ^hBu₄NF, THF, 23°C, 3 h. ⁱPh₃P/DEAD/PhCO₂H, PhH, 23°C, 1 h. ^j25% NaOMe/MeOH, THF, 23°C, 0.5 h.

trans-enal intermediate which underwent facile *in situ* Wittig olefination with the remaining 7-carbomethoxyhepta-3(Z)-en-1-ylidenetriphenylphosphorane¹² (7). Fluoride mediated deprotection of **5a** afforded methyl 12(R),20-dihydroxyeicosatetraenoate **5b**. Its enantiomer **5d** was secured via **5c** by inversion of the C(12)-alcohol using the Mitsunobu procedure,¹³ methanolysis of the derived benzoate, and desilylation as described above. Esters **5b,d** were converted to their free acids by saponification (NaOH, MeOH, 23°C, 4h), adjustment to pH 4.5, and extractive isolation.

Biological Evaluations

Canine polymorphonuclear leukocytes were incubated at 37°C with exogenous arachidonic acid and, following acidification, the incubate was extracted with EtOAc. Analysis of the organic soluble products by HPLC and GC/MS¹⁴ revealed **1** was a major oxygenated metabolite. Natural and synthetic **1** were tested for vasoactivity in canine coronary arteries by monitoring isometric tension of arterial rings suspended in a muscle bath containing Krebs' solution. Both enzymatically-derived and synthetic 12(S)-**1** relaxed the coronary artery. The latter showed a concentration dependent relaxation between 5 × 10⁻⁹ and 5 × 10⁻⁶ M in vessels precontracted with U46619 (20 nM). 12(S)-**1** caused a 48±8% relaxation at 5 × 10⁻⁶ M (p<0.01). In contrast, 12(R),20-DiHETE and 12(S)-HETE were without effect whereas 20-HETE contracted coronary arteries under basal tone. These studies indicate that

ω -hydroxylation of 12(S)-HETE or 12-hydroxylation of 20-HETE results in the formation of a vasodilator or, in the latter case, the conversion of a vasoconstrictor into a vasodilator. It would be of interest, consequently, to examine the potential role of 12,20-DiHETE in leukocyte-mediated myocardial reperfusion injury.¹⁵

Acknowledgment: Supported financially by the USPHS NIH (DK-38226, HL-17669, HL-51055) and the Robert A. Welch Foundation.

References and Notes

1. Hamberg, M.; Svensson, J.; Samuelsson, B. *Proc. Natl. Acad. Sci. USA* **1974**, *71*, 3824-3828. Malle, E.; Leis, H.J.; Karadi, I.; Kostner, G.M. *Int. J. Biochem.* **1987**, *19*, 1013-1022.
2. A 12(R)-lipoxygenase has been reported in the sea urchin: Hawkins, D.J.; Brash, A.R. *J. Biol. Chem.* **1987**, *262*, 7629-7634.
3. Capdevila, J.; Yadagiri, P.; Manna, S.; Falck, J.R. *Biochem. Biophys. Res. Comm.* **1986**, *141*, 1007-1011. Schwartzman, M.L.; Balazy, M.; Masferrer, J.; Abraham, N.G.; McGiff, J.C.; Murphy, R.C. *Proc. Natl. Acad. Sci. USA* **1987**, *84*, 8125-8129.
4. Review: Fretland, D.J.; Djuric, S.W. *Prost. Leuk. Essential Fatty Acids* **1989**, *38*, 215-228.
5. Wong, P.Y.-K.; Westlund, P.; Hamberg, M.; Granstrom, E.; Chao, P.H.-W.; Samuelsson, B. *J. Biol. Chem.* **1984**, *259*, 2683. Marcus, A.J.; Safier, L.B.; Ullman, H.L.; Broekman, M.J.; Islam, N.; Oglesby, T.D.; Gorman, R.R. *Proc. Natl. Acad. Sci. USA* **1984**, *81*, 903. Jajoo, H.K.; Capdevila, J.H.; Falck, J.R.; Bhatt, R.K.; Blair, I.A. *Biochim. Biophys. Acta* **1992**, *1123*, 110-116. Nishimura, M.; Schwartzman, M.L.; Falck, J.R.; Lumin, S.; Zirrollo, J.A.; Murphy, R.C. *Arch. Biochem. Biophys.* **1991**, *290*, 326-335. Asakura, T.; Shichi, H. *Biochem. Biophys. Res. Comm.* **1992**, *187*, 455-459.
6. Marcus, A.J. *Prog. Haemostasis Thromb.* **1986**, *8*, 127-142.
7. To date, only the 12(S),20-isomer has been synthesized: Manna, S.; Viala, J.; Yadagiri, P.; Falck, J.R. *Tetrahedron Lett.* **1986**, *27*, 2679-2682. Leblanc, Y.; Fitzsimmons, B.J.; Adams, J.; Perez, F.; Rokach, J. *J. Org. Chem.* **1986**, *51*, 789-793.
8. Dax, K.; Weidmann, H. *Adv. Carbohydr. Chem. Biochem.* **1976**, *33*, 189-234.
9. Robins, M.J.; Wilson, J.S. *J. Am. Chem. Soc.* **1981**, *103*, 932.
10. Spectral and physical data for **2c**: ¹H NMR (CDCl₃, 250 MHz) δ 2.03-2.13 (ddd, J=14.5, 6.04, and 4.88 Hz, 1H), 2.43 (d, J=14.5 Hz, 1H), 2.63 (dd, J=18.7 and 1.39 Hz, 1H), 2.81 (dd, J=18.7 and 7.4 Hz, 1H), 3.32 (s, 3H), 4.91 (br t, J=7.0 Hz, 1H), 5.09 (d, J=4.8 Hz, 1H), 5.11 (br t, J=5.7 Hz, 1H); ¹³C NMR (CDCl₃) δ 37.49, 39.48, 55.35, 78.16, 82.40, 105.46, 175.19; TLC (SiO₂) EtOAc/hexane (3:7), R_f = 0.17; Anal. calcd for C₇H₁₀O₄: C, 53.16; H, 6.37; Found: C, 53.22; H, 6.47. **3** (as mixture of anomers): ¹H NMR δ 2.01-2.53 (m, 4H), 2.76 (br s, 1H), 3.38 and 3.49 (s, total of 3H) 4.72-5.03 (m, 2H), 5.08-5.13 (m, 1H), 5.45-5.56 and 5.71-5.79 (m, total of 1H). **4a**: ¹H NMR δ 1.05 (s, 9H), 1.28-1.41 (m, 4H), 1.44-1.62 (m, 2H), 1.92-2.18 (m, 4H), 2.30-2.58 (m, 2H), 2.78 (d, J=11.4 Hz, 1H), 3.32 (s, 3H), 3.62 (t, J=6.4 Hz, 2H), 3.88 (dt, J=3.7 and 7.4 Hz, 1H), 4.06-4.17 (m, 1H), 4.99 (d, J=3.8 Hz, 1H), 5.39-5.57 (m, 2H), 7.28-7.38 (m, 6H), 7.59-7.65 (m, 4H); ¹³C NMR δ 19.22, 25.53, 26.86, 27.40, 28.89, 29.42, 32.50, 41.48, 54.69, 64.31, 71.25, 85.16, 105.32, 125.17, 127.56, 129.47, 132.23, 134.14, 135.56. **5b,d**: ¹H NMR δ 1.32-1.48 (m, 4H), 1.50-1.75 (m, 4H), 2.00-2.23

(m, 4H), 2.28-2.40 (m, 5H), 2.91 (br t, $J=7.5$ Hz, 2H), 3.58 (t, $J=7.2$ Hz, 2H) 3.62 (s, 3H), 4.00-4.34 (m, 1H), 5.33-5.62 (m, 5H) 5.72 (dd, $J=15.1$ and 5.3 Hz, 1H), 5.97 (br t, $J=11.0$ Hz, 1H), 6.62 (dd, $J=15.1$ and 11.0 Hz, 1H).

11. Manna, S.; Falck, J.R.; Chacos, N.; Capdevila, J. *Tetrahedron Lett.* **1983**, *24*, 33-36.
12. Lumin, S.; Falck, J.R.; Schwartzman, M.L. *Tetrahedron Lett.* **1991**, *32*, 2315-2318.
13. Review: Hughes, D.L. *Org. Reactions* **1992**, *42*, 335-656.
14. Rosolowsky, M.; Falck, J.R.; Willerson, J.T.; Campbell, W.B. *Circ. Res.* **1990**, *66*, 608-621.
15. Lucchesi, B.R. *Ann. Rev. Physiol.* **1990**, *52*, 561-576.

(Received in USA 5 July 1994; accepted 1 August 1994)